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(54) Title: REPLICATION INCOMPETENT HERPES VIRUSES FOR USE IN GENE THERAPY

(57) Abstract: Use of a replication incompetent herpes virus capable of delivering a gene to multiple connected sites within the ner-
vous system, which virus comprises: (a) a mutation which prevents or reduces the expression of at least two immediate early genes;
and (b) a heterologous gene operably linked to a promoter active during herpes virus latency; in the manufacture of a medicament for
the treatment of a central nervous system disorder, a method of determining whether a gene has an effect on a phenotype associated
with a central nervous system disorder and in a method of treatment of a disorder of the central nervous system.

REPLICATION INCOMPETENT HERPES VIRUSES FOR USE IN GENE THERAPY**Field of the Invention**

5 The present invention relates to replication incompetent herpes simplex viruses capable of efficiently transferring genes to multiple sites within the nervous system. It also relates to the use of such viruses in the study and treatment of diseases and conditions of the nervous system.

Background to the Invention

Herpes simplex viruses (HSV) 1 and 2 have often been suggested as a vector for gene delivery to the nervous system and also other cell types (for reviews see Coffin and Latchman 1995, Fink *et al.* 1996). As a vector HSV has a number of potential advantages in that it naturally enters latency in neurons, providing the possibility of long term gene expression, does not integrate into the host genome, preventing insertional mutagenesis (for example the activation of oncogenes or inactivation of tumour suppressor genes), can accept very large DNA insertions allowing the delivery of multiple genes, is easy to propagate, and can infect a wide variety of other cell types as well as neurons. HSV also has the unique ability among viruses currently under development as vectors in that it can be efficiently transported along nerves to the cell body (usually in the spine) by retrograde axonal transport following an initial peripheral infection. However, while this property of retrograde axonal transport of HSV vectors has been observed with replication competent vectors in the peripheral nervous system (PNS), it has not previously been exploited in vectors used in the central nervous system (CNS), probably due to limitations in the vectors which have previously been available.

While HSV1 is highly prevalent in the human population, in the vast majority of cases giving no obvious signs of disease, for use as a vector the virus must be disabled for safety and so as to minimise toxicity to target cells. Various strategies for disablement have been reported including the removal of genes which are unnecessary for growth in vitro but necessary for pathogenesis in vivo. Such genes include those encoding thymidine kinase (TK; Ho and Mokarski 1988),

ribonucleotide reductase (Goldstein and Weller 1988) and ICP34.5 (Coffin *et al.* 1995). However for minimal toxicity it has become apparent that expression of the regulatory immediate early genes ICPO, ICP4, ICP22 and ICP27, which are themselves cytotoxic, must be minimised (Johnson *et al.* 1994, Johnson *et al.* 1992, 5 Wu *et al.* 1996, Samaniego *et al.* 1998, Krisky *et al.* 1998). Such reductions in IE gene expression minimise transcription from the vast majority of the 80 or so other genes in the HSV genome. Removal of ICP4 or ICP27 completely prevents virus growth and so such deletions must be complemented in the cells used for virus propagation (e.g. Deluca *et al.* 1985). Deletion of ICP22 and/or ICPO, while these 10 genes are not absolutely essential for virus growth (Sacks and Shaffer 1987, Stow and Stow 1986, Post and Roizman 1981, Sears *et al.* 1985), reduces virus titre. Thus for the growth of HSV mutants with multiple IE gene deficiencies, cell lines must be produced which effectively complement deletions from the virus, and for effective growth of viruses with deletions in ICP4, ICP27, ICP22 and ICPO, all these 15 deficiencies would optimally need to be complemented. However as the IE proteins are highly cytotoxic (Johnson *et al.* 1994), IE gene expression in cell lines must be tightly regulated. This is usually achieved by the use of the homologous IE gene promoters which are relatively inactive in the absence of virus infection (e.g. E5 cells [ICP4], B130/2 cells [ICP27], E26 cells [ICP4 + ICP27], F06 cells [ICP4 + ICP27 + 20 ICPO]; Deluca and Schaffer 1987, Howard *et al.* 1998, Samaniego *et al.* 1995, Samaniego *et al.* 1997). This reduces the problem of IE protein cytotoxicity but still leaves an inherent problem in the generation of cells which are highly effective at complementing multiple IE gene deficiencies.

A second strategy to reduce IE gene expression, rather than deletion of the IE 25 genes themselves, is to include mutations in the gene encoding vmw65. vmw65 is a virion protein which transactivates IE promoters after virus infection (Batterson and Roizman 1983, Pellet *et al.* 1985), and while an essential structural protein, specific mutations abolish the trans-activating capability of the protein without affecting the structural integrity of the virus (Ace *et al.* 1989, Smiley and Duncan 1997). These 30 mutations vastly reduce IE gene expression although at high multiplicity or with the inclusion of hexamethylene bisacetamide (HMBA) in the media still allow efficient virus growth in culture (McFarlane *et al.* 1992).

Thus, for the construction of vector viruses the approach could also be taken of combining mutations in *vmw65*, which should reduce expression of all the IE genes, with deletion of ICP27 and/or ICP4, the two essential IE genes, giving viruses as above in which overall IE gene expression is minimised. This is the approach we
5 have taken to generate non-toxic HSV vectors in which IE gene expression has been minimised but which can still be grown in culture using cell lines containing the genes encoding ICP27, ICP4 and the equine herpes virus (EHV) homologue of *vmw65* (encoded by EHV gene 12; Thomas *et al.* 1999).

The development of HSV vectors has also required a second problem to be
10 overcome before they can be used to take advantage of the natural lifecycle of the virus in which a latent state is maintained in neurons. This problem results from the finding that in most cases promoters driving genes inserted into an HSV vector genome are rapidly transcriptionally inactivated as the virus enters latency, including the promoter which usually drives the expression of the only HSV transcripts present
15 during latency, the latency associated transcripts (LATs). To solve this problem, a number of approaches have been taken:

First, it was found that a Moloney murine leukaemia virus (MMLV) promoter linked to a fragment of the LAT promoter ('LAP1'; Goins *et al.* 1994) and inserted into the gene encoding glycoprotein C (gC) was able to drive expression
20 during latency, although neither LAP1 or the MMLV promoter alone, or a number of other promoters linked to LAP1 or alone allowed this to occur (Lokensgard *et al.* 1994). MMLV alone inserted in ICP4 (Dobson *et al.* 1990, Bloom *et al.* 1995) or in LAP1 (Carpenter and Stevens 1996) however was active, which may be speculated to be due to the proximity of these regions to the endogenous LATs rather than when
25 distant to the LAT region (in gC) as before. In other approaches it was found that LAP2 alone could give expression during latency when inserted in gC (Goins *et al.* 1994), but this expression was very weak, and that LAP2 linked to LAP1, like MMLV linked to LAP1, could also maintain latent gene expression when inserted in gC (Lokensgard 1997). Finally it was found that insertion of an internal ribosome
30 entry site (IRES) into the 2Kb LAT allowed expression of a downstream marker gene (with poly A site) during latency (Lachmann and Efstathiou 1997). However while the above approaches have demonstrated latent gene expression in the PNS, as

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yet latent gene expression in the brain has only been demonstrated in a very small number of transduced cells (Dobson *et al.* 1990, Bloom *et al.* 1995, Carpenter and Stevens 1996) and effective replication defective disabled HSV vectors for the long term gene transfer to the brain have not previously been available.

5 We have taken an alternative approach to the development of promoters allowing latent gene expression from within the context of an HSV vector where we have found that elements of the LAT region can be used to confer a long term activity onto promoters which are not usually active during latency by placing them downstream of a region designated LAT P2 (promoters inserted after HSV1 nt
10 120,219; Genbank file HEICG: W098/ 30707). For example a cytomegalovirus (CMV) IE promoter and some other promoters including a minimal neuron specific enolase (NSE) promoter and the Moloney murine leukaemia virus (MMLV) LTR promoter placed in such a context remain active during latency, even though when placed elsewhere latent gene expression with such promoters is generally very
15 inefficient.

Summary of the Invention

As discussed above, HSV has the unique property among viruses currently under development as vectors in that it can be efficiently transported along nerves
20 before entering latency in neuronal cell bodies. However it has not previously been found that this property can be exploited for gene delivery in the CNS using disabled herpes vectors, as replication incompetent, herpes vectors so far used in the CNS have not demonstrated such properties as gene delivery has always been limited to the site of inoculation. The present inventors have identified disabled, replication
25 incompetent herpes vectors which allow gene delivery to multiple regions within the brain following vector inoculation to only a single brain region. This results from highly efficient retrograde transport of the vectors to cell bodies at connected sites. The inventors have also identified the properties of the vectors which allows this to occur. Replication incompetent herpes vectors allowing such gene delivery to occur
30 combine the use of highly disabled replication incompetent vector backbones with promoter systems which are capable of directing gene expression in the long term. The surprising finding was made that vectors which were either less disabled than

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those provided by the invention (ie which potentially showed greater toxicity), or which contain promoter cassettes which are incapable of directing latent gene expression, do not allow such widespread gene delivery to occur, even in the short term. Replication incompetent herpes virus vectors as identified by the present
5 inventors may be used in methods of studying or treating disorders of the central nervous system. In particular such vectors may be used in methods of target validation, for example, by screening genes to identify genes important in a central nervous system disorder.

Accordingly, the present invention provides:

- 10 - a use of a replication incompetent herpes virus comprising:
 - (a) a mutation which prevents or reduces the expression of at least two immediate early genes; and
 - (b) a heterologous gene operably linked to a promoter active during herpes virus latency
- 15 in the manufacture of a medicament for use in treating or preventing a central nervous system disorder;
- a method of determining whether a gene has an effect on a phenotype associated with a central nervous system disorder or on a cell of the central nervous system which is relevant to a central nervous system disorder, which method
20 comprises:
 - (i) inoculating into a cell of the central nervous system *in vivo* or *in vitro* with a replication incompetent herpes virus comprising:
 - (a) a mutation which prevents or reduces the expression of at least
25 two immediate early genes; and
 - (b) a heterologous gene operably linked to a promoter active during herpes virus latency; and
 - (ii) monitoring a phenotype of said disorder or an effect of expression of said gene on said cell to determine thereby whether said gene has an effect on said cell or said phenotype;
- 30 - a method of screening genes implicated in a central nervous system disorder to identify a target for gene therapy or for small molecule modulators, which method

comprises a method of determining whether a gene influences a phenotype associated with a central nervous system disorder according to the invention;

- a method of treating a subject suffering from a disorder of the central nervous system, which method comprises administering a therapeutically effective amount of a replication incompetent herpes virus comprising:

(a) a mutation which prevents or reduces the expression of at least two immediate early genes; and

(b) a heterologous gene operably linked to a promoter active during herpes virus latency

to a subject suffering from a disorder of central nervous system;

The present invention is exemplified using vectors in which immediate early gene expression has been minimised by deletion of ICP27 and ICP4 and with a mutation minimising the transactivating capabilities of *vmw65*, although other similarly non-toxic vector backbones may be used. The invention is also exemplified using vectors in which the LAT region has been used to confer long term activity onto a promoter not usually active in the long term, although other promoter systems active during latency might be used, including those discussed above (Lokensgard *et al.* 1994, 1997, Goins *et al.* 1994, Lachmann and Efsthathiou 1997).

The term heterologous gene is intended to embrace any gene not found in the viral genome. The heterologous gene may be any allelic variant of a wild-type gene, or it may be a mutant gene. Heterologous genes are preferably operably linked to a control sequence permitting expression of said heterologous gene in a cell of the nervous system. Viruses of the invention may thus be used to deliver a heterologous gene/genes to cells of the nervous system where it will be expressed in multiple regions following inoculation to anatomically connected site(s).

Detailed Description of the Invention

Viruses

The present invention provides a method for identifying a gene that may be used as a target for therapeutic intervention in the treatment or prevention of a peripheral nervous system disorder. Such genes are identified by expressing a test gene in a cell of the central nervous system, preferably a neuronal cell, using a

replication incompetent herpes virus vector that expresses no or only minimal levels of the essential immediate early genes to deliver the gene to the cell and by determining whether expression of the gene affects a phenotype associated with a central nerve system disorder. A method of determining whether a gene influences a phenotype associated with a central nervous system disorder consists essentially of the following steps:

(i) inoculating the central nervous system *in vivo* or *in vitro* with a replication incompetent herpes virus comprising:

- (a) a mutation which prevents or reduces the expression of at least two immediate early genes; and
- (b) a heterologous gene operably linked to a promoter active during herpes virus latency; and

(ii) monitoring a phenotype of said disorder to determine thereby whether expression of said gene has an effect on said phenotype.

Typically the herpes virus is capable of delivering the heterologous gene to multiple connected sites within the nervous system such that the heterologous gene is expressed at one or more site distant from the site of inoculation. Preferably the heterologous gene is expressed at multiple connected sites in the central nervous system including the site where the virus was inoculated. Preferably, where the cell is in a neuronal culture the gene is expressed for extended periods such as from 1 month to 1 year, for example for 2, 3, 4, 5, or 6 months.

The present invention provides a method for screening genes to identify a gene which is important in a central nervous system disorder and which may be used as a target for identifying agents for treating the disorder.

The present invention also provides the use of a replication incompetent herpes virus vector that expresses no or only minimal levels of the essential immediate early genes as described herein in the manufacture of a medicament for the treatment of a central nervous system disorder. A method of treating of a subject suffering from a nervous system disorder by administering a therapeutically effective amount of a replication incompetent herpes virus vector as described herein to the central nervous system of a subject in need thereof is also provided by the present invention. The herpes virus vector is typically replication incompetent, does not

express significant levels of the immediate early genes ICP0, ICP4, ICP22 and ICP27 and infects at least one site in the CNS that is connected to but distant from the site of inoculation. For example, a virus inoculated into the striatum may be expressed in the substantia nigra.

5 A virus of the invention is capable of infecting a cell of the nervous system following inoculation, but is incapable of replicating in any cell unless at least one gene which has been rendered non-functional in the virus is also expressed in the cell, i.e. the virus is replication incompetent other than in the cells in which virus stocks are prepared.

10 By definition such viruses are attenuated such that they are replication incompetent. Viral regions altered for the purposes of such attenuation may be eliminated (completely or partly), made non-functional, or substituted by other sequences, in particular by a heterologous gene sequence. Attenuating mutations resulting in a replication incompetent virus have been described for all neurotrophic
15 herpes viruses so far described. A virus is replication incompetent if it is incapable of replicating in a cell which does not express at least one functional viral gene. Replication incompetent viruses are generally capable of replicating in cell lines which express a functional essential viral gene which compensates for the absence of a functional essential viral gene in the replication incompetent virus. Such cell lines
20 may be used to prepare viral stocks.

 Although the present invention has been exemplified using herpes simplex viruses, it will be understood that other neurotrophic viruses of the herpesviridae family may be modified such that they are replication incompetent and can achieve efficient gene delivery to multiple sites within the nervous system following
25 inoculation to a single site. In particular, such viruses may include varicella zoster virus, pseudo-rabies virus or bovine herpes viruses.

 When the virus of the invention is a herpes simplex virus, the virus may be derived from, for example, HSV1 or HSV2 strains, or derivatives thereof, preferably HSV1. Derivatives include inter-type recombinants containing DNA from HSV1 and
30 HSV2 strains. Such inter-type recombinants are described in the art, for example in Thompson *et al.* 1998 and Meignier *et al.* 1988. Derivatives preferably have at least 70% sequence homology to either the HSV1 or HSV2 genomes, more preferably at

least 80%, even more preferably at least 90 or 95%. Derivatives may share sequence homology or identity with a wild type HSV1 or HSV2 genome prior to mutation to produce a virus for use in a method of the invention. Preferably a derivative shares sequence homology or identity with a HSV1 or HSV2 viral strain falling within the
5 scope of the invention.

A derivative may have the sequence of a HSV1 or HSV2 genome modified by nucleotide substitutions, for example from 1, 2 or 3 to 10, 25, 50 or 100 substitutions. The HSV1 or HSV2 genome may alternatively or additionally be modified by one or more insertions and/or deletions and/or by an extension at either
10 or both ends.

For example the UWGCG Package provides the BESTFIT program which can be used to calculate homology (for example used on its default settings) (Devereux *et al.* (1984) *Nucleic Acids Research* **12**, p387-395). The PILEUP and BLAST algorithms can be used to calculate homology or line up sequences (typically
15 on their default settings), for example as described in Altschul (1993) *J. Mol. Evol.* **36**:290-300; Altschul *et al.* (1990) *J. Mol. Biol.* **215**:403-10.

Software for performing BLAST analyses is publicly available through the National Centre for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). This algorithm involves first identifying high scoring sequence pair (HSPs) by
20 identifying short words of length W in the query sequence that either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighbourhood word score threshold (Altschul *et al.*, 1990). These initial neighbourhood word hits act as seeds for initiating searches to find HSPs containing them. The word hits are extended in both
25 directions along each sequence for as far as the cumulative alignment score can be increased. Extensions for the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached.
30 The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLAST program uses as defaults a word length (W) of 11, the BLOSUM62 scoring matrix (see Henikoff and Henikoff (1992) *Proc. Natl. Acad.*

Sci. USA 89: 10915-10919) alignments (B) of 50, expectation (E) of 10, M=5, N=4, and a comparison of both strands.

The BLAST algorithm performs a statistical analysis of the similarity between two sequences; see e.g., Karlin and Altschul (1993) *Proc. Natl. Acad. Sci. USA* 90: 5873-5787. One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a sequence is considered similar to another sequence if the smallest sum probability in comparison of the first sequence to the second sequence is less than about 1, preferably less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

Viruses of the invention do not express, or express only very low levels of at least two immediate early genes. Very low levels are levels below the detection limit of a western blot using an antibody to a protein product of an immediate early gene. A virus may comprise any suitable mutation which minimises immediate early gene expression. Preferably a virus does not express, or expresses only very low levels of at least three immediate early genes. More preferably a virus does not express, or expresses only very low levels of at least four immediate early genes. The immediate early genes which must not be expressed or be expressed at only minimal levels are ICP0, ICP4, ICP22 and ICP27. The two immediate early genes may be ICP27 and ICP4, ICP27 and ICP0, ICP27 and ICP22, ICP4 and ICP0, ICP4 and ICP22 or ICP0 and ICP22.

Viruses may be functionally inactive for individual (e.g. ICP27 or ICP4) or multiple (ICP27 and/or ICP4 and/or ICP22 and/or ICP0 and/or ICP47) immediate early genes, or they may additionally contain a mutation in the gene encoding vmw65 reducing/preventing its transcriptional activation capability. Alternatively essential genes may be deleted which are not immediate early genes, either alone, in any combination, or in combination also with the inactivation of an immediate early gene or genes or vmw65. Examples of such viruses include viruses reported by Samaniego *et al.* 1998, Krisky *et al.* 1998, or Thomas *et al.* 1999. Particularly preferred embodiments include viruses in which immediate early gene expression has been prevented or minimised such as those with inactivating mutations in or

deletions of the genes encoding ICP4, ICP27, ICP0 and ICP22, or alternatively with inactivating mutations in or deletions of ICP27 and/or ICP4 with a mutation in vmw65 reducing or preventing its trans-activating activity.

The terminology used in describing the various HSV genes is as found in
5 Coffin and Latchman, 1996.

Production of Viruses

When the herpes simplex viruses of the invention lack a particular functional essential gene, for example a gene encoding ICP4 or ICP27, the virus of
10 the invention is propagated on a cell line expressing that essential gene. For example, when the virus lacks a functional ICP27 gene, the virus may be propagated on V27 cells (Rice and Knipe, 1990), 2-2 cells (Smith *et al.*, 1992) or B130/2 cells (Howard *et al.* 1998). When the virus lacks a functional ICP4 gene the virus may be propagated on a cell line expressing ICP4, for example E5 cells (DeLuca *et al.*,
15 1985). When the virus lacks a functional ICP4 gene and a functional ICP27 gene the virus is propagated on a cell line expressing both ICP4 and ICP27 (such as E26 cells; Samaniego *et al.*, 1995), and when the virus additionally lacks a functional vmw65 gene the virus may be propagated on a cell line also containing a non-HSV homologue of vmw65 (e.g. from equine herpes virus as in Thomas *et al.* 1999).
20 Mutations to vmw65 may also be partially compensated for by inclusion of hexamethylene bisacetamide (HMBA) in the media used for virus growth (MacFarlane *et al.* 1992).

ICP27-expressing cell lines can be produced by co-transfecting mammalian cells, for example the Vero or BHK cells, with a vector, preferably a plasmid vector,
25 comprising a functional HSV ICP27 gene capable of being expressed in said cells. and a vector, preferably a plasmid vector, encoding a selectable marker, for example neomycin resistance. Clones possessing the selectable marker are then screened further to determine which clones also express functional ICP27. for example on the basis of their ability to support the growth of ICP27- HSV strains, using methods
30 known to those skilled in the art (for example as described in Rice and Knipe, 1990).

Cell lines which do not allow reversion of an ICP27- mutant HSV strain to a strain with functional ICP27 are produced as described above, ensuring that the

vector comprising a functional ICP27 gene does not contain sequences that overlap with (i.e. are homologous to) sequences remaining in the ICP27- mutant virus.

Where HSV strains of the invention comprise inactivating modifications in other essential genes, for example ICP4, complementing cell lines will comprise a functional HSV gene which complements the modified essential gene in the same manner as described for ICP27. For example in the case of HSV strains comprising mutations in both ICP27 and ICP4, a cell line expressing both ICP27 and ICP4 is used such as E26 cells (Samaniego *et al.*, 1995). HSV strains expressing other essential genes can be constructed in a similar manner to that described for ICP27. Here again, if it is ensured there is no sequence overlap between the remaining virus DNA and that inserted into the cell line for virus growth, the possibility of reversion of the virus to a less disabled form during growth will be minimised.

Methods of mutation

The various viral genes referred to may be rendered functionally inactive by any suitable technique. For example, they may be rendered functionally inactive by deletions, substitutions or insertions, preferably by deletion. Deletions may remove portions of the genes or the entire gene. For example, deletion of only one nucleotide may be made, resulting in a frame shift. However, preferably larger deletions are made, for example at least 25%, more preferably at least 50% of the total coding and non-coding sequence (or alternatively, in absolute terms, at least 10 nucleotides, more preferably at least 100 nucleotides, most preferably, at least 1000 nucleotides). It is particularly preferred to remove the entire gene and some of the flanking sequences. Inserted sequences may include the heterologous genes described below. In particular, it is preferred to insert the heterologous gene into ICP27 or ICP4. In the case of the *vmw65* gene, the entire gene is not deleted since it encodes an essential structural protein, but a small inactivating mutation is made which abolishes the ability of VMW65 to activate transcriptionally IE genes (e.g. as in Ace *et al.*, 1989 or Smiley *et al.* 1997).

Mutations are made in the herpes viruses by homologous recombination methods well known to those skilled in the art. For example, HSV genomic DNA is transfected together with a vector, preferably a plasmid vector, comprising the

mutated sequence flanked by homologous HSV sequences. The mutated sequence may comprise deletions, insertions or substitutions, all of which may be constructed by routine techniques. Insertions may include selectable marker genes, for example lacZ or GFP, for screening recombinant viruses by, for example, b-galactosidase
5 activity or fluorescence.

Heterologous genes and promoters

The viruses for use in the invention are typically modified to carry a heterologous gene. The term "heterologous gene" encompasses any gene. A
10 heterologous gene may be a test gene for use in a method of target validation or a gene of therapeutic use for use in a method of treatment or in the manufacture of a medicament. Although a heterologous gene is typically a gene not present in the genome of a herpes virus, herpes gene/genes may be used provided that the coding sequence is not operably linked to the viral control sequences with which it is
15 naturally associated.

The heterologous gene may be any allelic variant of a wild-type gene, or it may be a mutant gene. The term "gene" is intended to cover nucleic acid sequences which are capable of being at least transcribed. Thus, sequences encoding mRNA, tRNA and rRNA are included within this definition. However, the present invention
20 is concerned with the expression of polypeptides or antisense RNA rather than tRNA and rRNA. The term "gene" covers a polynucleotide comprising the coding sequence of a polypeptide. The polynucleotide may include one or more introns, for example may comprise genomic DNA. Preferably the polynucleotide comprises cDNA. Sequences encoding mRNA or antisense RNA will optionally include some
25 or all of 5' and/or 3' transcribed but untranslated flanking sequences naturally, or otherwise, associated with the translated coding sequence. It may optionally further include the associated transcriptional control sequences normally associated with the transcribed sequences, for example transcriptional stop signals, polyadenylation sites and downstream enhancer elements.

30 A heterologous gene may be inserted into the viral genome by homologous recombination of HSV strains with, for example, plasmid vectors carrying the heterologous gene flanked by HSV sequences. The heterologous gene may be

introduced into a suitable plasmid vector comprising herpes viral sequences using cloning techniques well-known in the art. The heterologous gene may be inserted into the viral genome at any location provided that the virus can still be propagated. It is preferred that the heterologous gene is inserted into an essential gene.

- 5 Heterologous genes may be inserted at multiple sites within the virus genome.

The transcribed sequence of a heterologous gene is preferably operably linked to a control sequence permitting expression of the heterologous gene in a mammalian cell, preferably a cell of the peripheral nervous system. The term "operably linked" refers to a juxtaposition wherein the components described are in a relationship
10 permitting them to function in their intended manner. A control sequence "operably linked" to a coding sequence is ligated in such a way that expression of the coding sequence is achieved under conditions compatible with the control sequence.

The heterologous gene is under the control of a promoter active during herpes virus latency. Such promoters typically comprise fragments of the latency associated
15 transcript (LAT) region of the virus or are chimaeric promoters comprising fragments of the LAT region together with a non-LAT promoter. Such latently active promoters may drive the expression of endogenous gene or genes from within the LAT region of the virus, or when inserted at an ectopic, non-LAT site within the HSV genome.

20 The control sequence comprises a promoter allowing expression of the heterologous gene and a signal for termination of transcription. The promoter is selected from promoters which are functional during herpes latency in mammalian, preferably rat, mouse, guinea pig, ferret, primate or human, cells of the central nervous system. The promoter/promoters may be derived from promoter sequences
25 of eukaryotic genes. For example, promoters may be derived from the genome of a cell in which expression of the heterologous gene is to occur, preferably a mammalian, preferably human cell of the peripheral nervous system. With respect to eukaryotic promoters, they may be promoters that function in a ubiquitous manner (such as promoters of β -actin, tubulin) or, alternatively, a tissue-specific manner,
30 such as the neuron-specific enolase (NSE) promoter. They may also be promoters that respond to specific stimuli, for example promoters that bind steroid hormone receptors. Viral promoters may also be used, for example the Moloney murine

leukemia virus long terminal repeat (MMLV LTR) promoter or other retroviral promoters, the human or mouse cytomegalovirus (CMV) IE promoter, or promoters of herpes virus gene including those driving expression of the latency associated transcripts.

5 Expression cassettes and other suitable constructs comprising a heterologous gene and control sequences can be made using routine cloning techniques known to persons skilled in the art (see, for example, Sambrook *et al.*, 1989, Molecular Cloning – A Laboratory Manual; Cold Spring Harbor Press).

 It may also be advantageous for the promoters to be inducible so that the
10 levels of expression of the heterologous gene can be regulated during the life-time of the cell. Inducible means that the levels of expression obtained using the promoter can be regulated. For example, in a preferred embodiment where more than one heterologous gene is inserted into the HSV genome, one promoter would comprise a promoter responsive to the tet repressor/VP16 transcriptional activator fusion protein
15 previously reported (Gossen and Bujard, 1992, Goßsen *et al.*, 1995), and driving the heterologous gene the expression of which is to be regulated. The second promoter would comprise a strong promoter (e.g. the CMV IE promoter) driving the expression of the tet repressor/VP16 fusion protein. Thus in this example expression of the first heterologous gene would depend on the presence or absence of
20 tetracycline.

 In the current invention, a promoter, or more than one promoter, is used which allows gene expression during herpes virus latency. Such promoters may be derived from the LAT region (e.g. LAP2; Goins *et al.* 1994) and may be chimaeric promoters containing sequences from the LAT region fused to a non-LAT promoter
25 or other elements (e.g. Lachmann and Efsthathiou, 1997). These promoters may function from within the endogenous LAT region of the virus or when inserted at a heterologous site in the virus genome. In particularly preferred embodiments chimaeric promoters consisting of a non-LAT promoters such as the CMV, MMLV or NSE promoters, or fragments thereof, linked to LAT-derived sequences, including
30 those present in LAP1 (Goins *et al.* 1994), LAP2 (Goins *et al.* 1994) and/or LAT. The LAT P2 region is here defined as HSV1 nucleotides 118866-120219 of HSV strain 17+ (GenBank HE1CG: from PstI-BstXI sites), and includes fragments or

derivatives of this region, including homologous regions of other HSV1 strains and of HSV2 strains, which are capable of providing a long-term expression capability to promoters to which they are linked. Such chimaeric promoters may also allow the expression of multiple heterologous genes by placing pairs of promoters away from a central LAT-derived element such that two genes may be expressed from a single expression cassette (e.g. as in Thomas *et al.* 1999).

Heterologous genes in viruses for therapeutic uses will typically encode polypeptides of therapeutic use. For example, genes which may prevent further degeneration of the cells affected in a particular disease, stimulate new cells to be produced, or affect the metabolic functioning (e.g. by the delivery of genes encoding enzymes) of the cell. Combinations of such factors might also be delivered using a single vector virus. Heterologous genes may be wild-type genes or may be mutated to enhance the function of the gene or to inhibit function of the gene. For example, where an endogenous gene is over expressed in a central nervous system disorder, a non-functional gene which disrupts the function of the endogenous gene may be useful in treating the disorder.

In particular, the heterologous gene/genes may encode a polypeptide capable of preventing, reducing or reversing cellular degeneration in degenerative diseases such as Parkinson's disease or Alzheimer's disease, or in increasing the activity of the remaining cells in such diseases. Such genes include neurotrophic factors such as GDNF and related proteins, or enzymes such as tyrosine hydroxylase or GTP-cyclohydrolase each of which may be of benefit in the treatment of Parkinson's disease, or genes encoding proteins such as Sonic Hedgehog or Nurr-1. Other examples of potentially beneficial genes include those which are deficient in metabolic diseases such as Tay-Sachs or mucopolysaccharide diseases or other diseases for which a genetic defect is known.

Suitable heterologous genes include growth factors such as NT-3, Reg-2, CNTF, GDNF, BDNF and NGF and factors of unknown function. The present invention may be used to attempt to identify the function of such factors, for example by determining the effect of the factor of unknown function on a cell of the central nervous system. Genes encoding factors of unknown function may typically have been implicated in a central nervous system disorder.

Heterologous genes may also include marker genes (for example encoding β -galactosidase or green fluorescent protein or other fluorescent proteins) or genes whose products regulate the expression of other genes (for example, transcriptional regulatory factors including the tet repressor/vmw65 transcriptional activator fusion protein described above).

Target validation, gene therapy, and other therapeutic applications may well require the administration of multiple genes. The expression of multiple genes may be advantageous for the treatment of a variety of conditions. Herpes viruses are uniquely appropriate as they do not have the limited packaging capabilities of other viral vector systems. Thus multiple heterologous genes can be accommodated within its genome. There are, for example, at least two ways in which this could be achieved. For example, more than one heterologous gene and associated control sequences could be introduced into a particular HSV strain either at a single site or at multiple sites in the virus genome. It would also be possible to use pairs of promoters (the same or different promoters) facing in opposite orientations away from each other, these promoters each driving the expression of a heterologous gene (the same or different heterologous gene) as described above.

A virus may comprise one, two, three, four or more heterologous genes. Where the gene is a test gene for use in a method of target validation, the virus typically comprises one heterologous gene. The virus may comprise one, two or more copies of a heterologous gene. A virus for use in a method of target validation may comprise more than one heterologous test gene. This will enable several genes, for example from 2 to 20 genes, preferably from 2 to 10 genes, for example, 4, 6 or 8 genes, to be screened simultaneously. If an effect of a test gene is observed, further viruses carrying fewer test genes, for example one or two test genes, may be used to determine which of the genes is responsible for the observed effect.

Heterologous genes for use in target validation methods will typically be implicated in a central nervous system disorder. For example, a gene may be implicated in a disorder due to its presence on region of a chromosome linked to the disorder. Alternatively a gene may be implicated in a central nervous system disorder if it is known to be expressed in the central nervous system. A gene implicated in a central nervous system disorder may be upregulated or down-

regulated in a tissue in a natural, artificial or induced disease state. Other suitable test genes include genes that encode proteins which are involved in nerve cell growth and/or regeneration such as neuronal growth factors.

5 Monitoring Phenotype

As used herein the term "phenotype" is used to refer to any physical manifestation of a disorder of the central nervous system. The term "phenotype" is thus intended to include changes at the molecular, cellular, tissue, cognitive and behavioural levels. Damage and trauma to the central nervous system, for example
10 as a result of physical and chemical injury is also included within the term "phenotype".

Any suitable phenotype associated with a central nervous system disorder may be monitored. For example, the motor control of an animal may be monitored using any suitable assay for monitoring the motor activity or behavior of an animal in
15 response to a suitable stimulus. The test animal may be a rodent, primate or other laboratory animal. A test gene which influences the phenotype will typically enhance or diminish the response of the animal to a given stimulus. Control responses may be determined by testing an animal prior to inoculation with the virus or by testing one or more control animals. A control animal is an animal which has
20 not been inoculated with a virus or which has been inoculated with a control virus which does not express a heterologous gene or which expresses a heterologous gene that does not affect the phenotype of interest. Cognitive function of an animal may also be monitored. Assays for monitoring motor activity and cognitive function are known in the art.

25 A phenotype associated with a central nervous system disorder may also be monitored at the cellular or molecular level. This is particularly important when monitoring the effect of a test gene *in vitro*. For example, the cellular response to the application of a chemical stimulus such a neurotransmitter or agonist or antagonist of a neurotransmitter receptor may be monitored. This may be achieved using
30 electrophysiological techniques. Alternatively any changes in gene expression within a central nervous system cell in response to the test gene may be monitored.

A phenotype may be the susceptibility of an animal to a disease. A method of the invention may therefore be a method of determining the role of a gene in the

susceptibility of an animal to a nervous system disorder. For example, following inoculation of a test animal with a test gene, a factor associated with a nervous system disorder may be administered to the animal, or the animal may be exposed to such a factor, and the onset of symptoms of the disorder may be monitored and
5 compared to the onset of symptoms in a control animal. Typically, a factor associated with a nervous system disorder is thought to be a causative factor in the onset of the disorder. Alternatively, onset or development of a disease may be monitored over time in the absence of a specific factor. For example, a test gene may promote or delay the onset of a neurological disorder.

10 A test gene may be suitable as a target for gene therapy or for small molecule modulators useful in the treatment of a central nervous system disorder if it influences a phenotype associated with the disorder. A gene identified as a target using a method of the invention may be used in a method of screening for agents useful in the treatment of a central nervous system disorder if it influences a
15 phenotype associated with the disorder. Generally a test gene may be considered to influence a phenotype if it inhibits or enhances the phenotype, for example expression of a phenotype may be increased or decreased by at least 5%, for example by at least 10%, at least 15%, at least 20% or at least 25%, preferably by at least 30%, for example at least 40% or at least 50%, more preferably by at least 70%, for
20 example, at least 80% or at least 90% compared to controls.

Introducing virus

A replication incompetent herpes virus as described herein may be introduced into the central nervous system by any suitable means. For example, by injection
25 into the brain, spinal cord or ventricles within the brain. A replication incompetent herpes virus as described herein is preferably introduced into one or more sites within the brain, for example into the striatum, spinal cord, cortex, cerebellum, hippocampus or brain stem. The expression of a heterologous reporter gene in the virus may be monitored at the site of delivery and at remote sites within the central
30 nervous system to determine the efficiency of gene expression. Preferably, the heterologous gene is expressed at the site of delivery and at least one connected site.

Therapeutic uses

Viruses of the invention may be used in methods of therapy in the treatment of nervous system disorders. As used herein, the term "disorder" is intended to include any condition of the nervous system which affects the normal functioning of the nervous system. "Conditions", "diseases" and "injury" to the central nervous system are thus encompassed by the term "disorder". The term "disorder" also includes animal models of peripheral nervous system disorders. In particular, viruses of the invention may be used for the treatment of neurodegenerative diseases such as Parkinson's disease, stroke or Alzheimers disease, or in single gene defect diseases such as mucopolysaccharide diseases or Tay-Sachs disease.

Replication incompetent herpes viruses capable of infecting cells of the central nervous system and expressing a heterologous gene at a site connected to but distinct from the site of infection as described herein may be used in methods of therapy. In particular, viruses of the invention may be used for the treatment of neurodegenerative diseases.

The herpes viruses of the present invention may thus be used to deliver therapeutic genes to a human or animal in need of treatment. Delivery of therapeutic genes using the herpes viruses of the invention may be used to treat central nervous system disorders, for example diseases such as Alzheimer's disease or Parkinson's disease, or to improve cognitive function, for example following stroke injury. Generally, the condition of a patient suffering from a central nervous system disorder is improved by administration of the virus. Administration of the herpes virus vector to a subject suffering from disorder of the central nervous system typically alleviates the symptoms of the disorder an/or prevents the progression of the disorder.

One method of administering therapy involves inserting a therapeutic gene into the genome of the herpes virus of the invention, as described above, and then combining the resultant recombinant virus, as active ingredient, with a pharmaceutically acceptable carrier or diluent to produce a pharmaceutical composition. Suitable carriers and diluents include isotonic saline solutions, for example phosphate-buffered saline.

Gene delivery to cells of the peripheral nervous system may be carried out by direct injection of the vector composition into a single site, or into two or more sites,

of the central nervous system of a human or animal. The amount of virus administered is in the range of from 10^4 to 10^{10} pfu, preferably from 10^5 to 10^8 pfu, more preferably about 10^6 to 10^8 pfu. When injected, typically from 1-200 μ l preferably from 1 to 10 μ l of virus suspension, depending on the species, in a
5 pharmaceutically acceptable suitable carrier or diluent, is administered.

The routes of administration and dosages described are intended only as a guide since a skilled practitioner will be able to determine readily the optimum route of administration and dosage. The dose of a modulator may be determined according to various parameters, especially according to the substance used; the age, weight
10 and condition of the patient to be treated; the route of administration; and the required regiment. A physician will be able to determine the required dosage for any particular patient.

The herpes viruses of the present invention may thus be used to deliver therapeutic genes to a human or animal in need of treatment. Delivery of therapeutic
15 genes using the herpes viruses of the invention may be used to treat for example Parkinson's disease or Alzheimer's disease or other neurodegenerative conditions.

One method of administering therapy involves inserting the therapeutic gene/genes into the genome of the herpes virus of the invention, as described above, and then combining the resultant recombinant virus with a pharmaceutically
20 acceptable carrier or diluent to produce a pharmaceutical composition. Suitable carriers and diluents include isotonic saline solutions, for example phosphate-buffered saline.

Gene delivery to cells of the nervous system may then be carried out following direct injection of the vector composition into a site or into two or more
25 sites within the brain of a human or animal. The amount of virus administered is in the range of from 10^4 to 10^4 pfu, preferably from 10^5 to 10^8 pfu or 10^6 to 10^8 pfu, more preferably about 10^6 to 10^7 pfu. When injected, typically from 1 to 200 μ l, preferably from 1 to 10 μ l, of virus suspension in a pharmaceutically acceptable suitable carrier or diluent, depending on the species and site of inoculation, is
30 administered.

The routes of administration and dosages described are intended only as a guide since a skilled practitioner will be able to determine readily the optimum route

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of administration and dosage for any particular patient depending on , for example, the age, weight and condition of the patient.

The invention will be described with reference to the following Examples,
5 which are intended to be illustrative only and not limiting.

EXAMPLES

Materials and Methods

5 Construction and Growth of Viral Strains

Replication incompetent virus strains are derived from HSV1 strain 17+, the nucleotide sequence of which is deposited in GenBank (Accession No. HE1CG). These can be propagated on 27/12/M:4 cells which are BHK cells containing the HSV ICP27 and ICP4-encoding genes and equine herpes virus gene 12 (Thomas *et al.*, 1999). These cells allow efficient growth of viruses with mutations in ICP4,
10 ICP27 and with mutations in vmw65 reducing its transactivating activity. Alternatively B130/2 cells which complement mutations to only ICP27 (Howard *et al.* 1998) are used for the growth of viruses disabled by the mutation ICP27 alone. These cell lines only complement the genes which have been deleted and there is no
15 sequence overlap between the viral and cellular genomes.

The pR20.5 and pR19 promoter cassettes are shown in Figure 1. The pR20.5 cassette was inserted into a plasmid allowing insertional inactivation of UL41, the gene encoding vhs (insertion at the unique *Nru*I site at the nucleotide 91854 in UL 41). The pR19lacZ and pR19GFP cassettes used here (see Figure 1) were
20 recombined into the endogenous LAT regions between the two *Bst*XI sites (nt 120220 and 120408). HSV1 nucleotide numbers refer to GenBank file HE1CG.

The current invention was exemplified using virus strain 1764/27-/4-/pRI9lacZ. The term 1764 describes a virus with the *in*1814 mutation in the gene encoding VMW65 and with the genes encoding ICP34.5 and ORF P completely
25 deleted (between nucleotides nt 124945-125723). The term 27- refers to the deletion of nucleotides 113273-116869, which contain the genes UL54, 55 and 56. UL54 is the gene encoding the essential IE gene ICP27 and UL55 and 56 are both non-essential genes. Hence, 1764/27-/4- virus is deleted for ICP34.5, ICP27 and ICP4 with an inactivating mutation in vmw65 as previously described for virus strain
30 1764/27-/4-/PR20.5 (Thomas *et al.* 1999). This virus however contains a CMV promoter (from pcDNA3; Invitrogen)/lacZ (pCH I 10; Pharmacia)/poly A (from pcDNA3) cassette inserted into both copies of the LAT region between the *Bst*XI

sites at nts 120,219 and 120,413 rather than the insertion into ICP4 described in Thomas *et al.* 1999. Before insertion of the CMV/lacZ cassette in 1764/27-/4-/pR191acZ the lacZ/GFP insertion into ICP4 in 1764/27-/4-/PR20.5 was removed by recombination with empty ICP4 flanking regions and selection of virus plaques which did not express lacZ or GFP giving virus strain 1764/27-/4-/w. The pR191acZ cassette is capable of gene expression during HSV latency as described in W0 98/ 30707.

Virus strain 17+/pRI91acZ contains an identical insertion into the LAT region as in virus strain 1764/27-/4-/pRI91acZ, is disabled only by the deletion of ICP27, and has been described before (Wagstaff *et al.* 1998). Virus strain 1764/27-/4-/CMVlacZ/US5 contains a CMV/lacZ/polyA insertion into the unique SacI site in the non-essential US5 gene of 1764/27-/4-/w. The CMV promoter in such a context is not active during latency.

Virus strains 1764/27- and 1764/27-/4- were also deleted for the endogenous LAT P2 regions (between nucleotides 118768 and 120470, *DdeI* to *HpaI*) in order to prevent the recombinational instability which has otherwise been found to occur following insertion of LAT P2 containing expression cassettes outside of the LAT region.

The viral vector backbones used in the Examples are summarised in Figure 1a and the promoter cassettes used in the Examples are shown in Figure 1b.

Example 1: Replication incompetent HSV vectors containing a long-term promoter and deleted for ICP27 do not allow widespread gene delivery in the brain following inoculation at a single site.

5 μ l of virus strain 17+/pRI91acZ at a titre of 1×10^8 pfu/ml was inoculated into the striatum of 200-220g female Lewis rats using stereotaxic procedures and a glass micropipette for inoculation. Virus suspension was administered at a rate of 0.5 μ l/min. Pairs of animals were perfusion fixed at either 3 days, 1 week or 1 month post-inoculation, and sectioned and stained with X-gal as previously described (Howard *et al.* 1998). Brain sections showed robust X-gal staining in the striatum at 3 days post-inoculation which had reduced to some extent by 1 week and which was evident in only small numbers of cells by 1 month. However there was no significant X-gal staining evident at sites in the brain connected to the striatum such as the

substantia nigra, from which dopaminergic neurons project, at any of the time points tested. Therefore using a replication incompetent virus backbone which still allows significant immediate early gene expression (ICP27 deleted; Thomas *et al.* 1999) and a promoter which does otherwise allow gene expression during herpes latency gene delivery is only observed at the inoculated site within the brain and not also at connected sites such as the substantia nigra (other than a small number of cells) where significant gene expression might have been expected had efficient retrograde transport followed by gene expression occurred.

- 10 **Example 2: Replication incompetent HSV vectors containing a short-term promoter, deleted for ICP27 and ICP4 and with an inactivating mutation in vmw65, do not allow widespread gene delivery in the brain following inoculation at a single site.**

5µl of virus strain 1764/27-/4-/CMVlacZ/US5 at a titre of 1×10^8 pfu/ml was inoculated into the striatum of 200-220g female Lewis rats using stereotaxic procedures as in Example 1. Pairs of animals were again perfusion fixed at either 3 days, 1 week or 1 month post-inoculation, and sectioned and stained with X-gal. Brain sections again showed robust X-gal staining in the striatum at 3 days post-inoculation which had reduced by 1 week and but which in this case was undetectable by 1 month post-inoculation. There was no X-gal staining evident at sites in the brain connected to the striatum at any of the time points tested. Therefore using a replication incompetent virus backbone in which immediate early gene expression has been minimised (ICP4 and ICP27 deleted, vmw65 mutation reducing transactivation; Thomas *et al.* 1999) and a promoter which does not allow gene expression during herpes latency, gene delivery is again only observed at the inoculated site within the brain and not also at connected sites such as the substantia nigra where gene expression might have been expected had retrograde transport followed by gene expression occurred.

- 30 **Example 3: Replication incompetent HSV vectors containing a long-term promoter, deleted for ICP27 and ICP4 and with an inactivating mutation**

in vmw65 allow gene delivery to multiple sites in the brain following inoculation to only a single site.

Injections into the rat striatum, spinal cord or superior colliculus were carried out with the 1764/27-/4-/pR19lacZ. 5µl of virus strain 1764/27-/4-/pR19lacZ at a titre of 1×10^8 pfu/ml was inoculated into the striatum of 200-220g female Lewis rats as above and brains removed and sectioned at time points as before. Brain sections again showed high efficiency gene delivery in the striatum at 3 days post-inoculation with large numbers of X-gal staining cells also evident at 1 week and 1 month after inoculation. However, unlike in the previous Examples, significant X-gal staining was observed in the substantia nigra. Therefore using both a replication incompetent virus backbone in which IE gene expression has been minimised and a promoter active during virus latency, gene delivery is observed at both the inoculated site within the brain, in this case the striatum, and also at connected sites such as in this case the substantia nigra. This results from highly efficient retrograde transport following inoculation in the striatum allowing gene expression in neuronal cell bodies in the substantia nigra from which they project.

Adult Lewis rats were also stereotactically injected with 2.5×10^5 pfu of 1764/27-/4-/pR19lacZ at the C6 level of the spinal cord or into the superior colliculus and thalamus. Three weeks after injection, the animals were perfusion fixed and relevant areas of the nervous system were sectioned and stained for lacZ expression.

In each case, the virus gave high level gene expression at the site of injection and was also efficiently transported from the site of injection to the cell bodies at connected sites where gene expression occurred. This was clearly demonstrated by the transport of the disabled virus from the striatum to the substantia nigra, from the superior colliculus via the optic nerve to the retinal ganglion cells, and from the C6 level of the spinal cord to DRGs, the brainstem and areas of the hind and midbrain.

Example 4: Virus strain 1764/27-/4- does not express significant amounts of any immediate early gene.

To test the levels of residual IE gene expression of a virus deleted for ICP27 and/or ICP4 and with VMW65 inactivated, non-complementing BHK cells were infected at MOIs of 10, 5 and 1 with 17+27-, 1764 27- and 1764 27- 4- viruses.

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Complementing 27/12/M:4 cells were infected with the same viruses at an MOI of 1 as positive control. Cells were harvested 48 hours post infection and analyzed for the expression of ICP0, ICP22 and ICP47 by western blotting. Samples for western blot analysis were prepared by standard techniques. Extract from approximately 10^5 cells was loaded per lane of an SDS-polyacrylamide gel. Proteins were transferred to nitrocellulose and probed with appropriate antibodies by standard techniques. Antibody against HSV1 ICP0 was purchased from ABI. Antibodies to HSV1 ICP22, ICP6 and ICP47 were provided by Bernard Roizman, Barklie Clements and David Johnson respectively. Detection was performed using ECL chemiluminescence (Amersham).

The 17+27- virus expressed significant amounts of all three of the IE genes which have not been deleted (ICP0, ICP22 and ICP47). The subsequent inactivation of VMW65 and deletion of ICP34.5 (the 1764 27- virus) leads to reduced but still significant levels of ICP0, ICP22 and ICP47 expression. This suggests that the inactivating mutation in VMW65 in the context of an ICP27 deleted virus is not sufficient to completely prevent the transactivation of the remaining IE gene promoters. However, prevention of transactivation of the other IE gene promoters can be achieved by the additional deletion of ICP4 (the 1764 27- 4- virus). The western blots for ICP0, ICP22 and ICP47 reveal that there is no detectable expression of any of these IE genes from this virus, even at high multiplicity of infection.

The 1764 27- 4- virus does not therefore express significant amounts of any of the IE genes. The essential IE genes ICP4 and ICP27 have been completely deleted from the viral backbone and the remaining IE genes ICP0, ICP22 and ICP47 are not expressed. This lack of IE gene expression requires the combination of VMW65 inactivation and ICP4/ICP27 deletion and provides a level of disablement to the virus similar to that resulting from the individual deletion of each of the IE genes (Samaniego *et al.*, 1998).

Example 5: Virus strain 1764/27-/4-pR20.5 simultaneously expresses high levels of two exogenous genes

It has been previously reported that in the absence of IE gene products, the expression of inserted genes is undetectable in the majority of transduced cells even at short times post infection (Saminego *et al.* 1998). It has been proposed that this absence of transgene expression is attributable to an active repression of both HSV
5 and non-HSV promoters by a cellular factor, but that this effect is masked in less disabled viruses by the presence of the potent viral transactivators VMW65, ICP0 and ICP4 (Preston and Nicholl 1997). Non-complementing BHK cells were infected at an MOI of 10 and visualized by fluorescence microscopy or processed to detect lacZ expression 48 hours post infection. Abundant transgene expression from the
10 1764/27-/4-pR20.5 virus was observed 48 hours post infection even though IE expression levels were minimal.

Example 6: Exogenous gene expression from cassettes inserted into virus strain 1764/27-/4- is dose dependent in non-complementing cells.

15 The expression level of ICP22 and ICP47 from the 1764/27- virus in non-complementing cells was not dose dependent, the peak of expression occurring at an MOI of 1. This suggests that with this intermediately disabled virus, interactions between the virus genome and remaining viral factors are affecting virus gene expression. In order to examine this further, non-complementing BHK cells were
20 infected with a selection of disabled viruses over a wider range of MOIs and the samples harvested 48 hours post infection. Western blots were then probed for the expression of ICP22 and ICP47 as before. This experiment showed that in the case of the 17+/27- virus, ICP22/47 expression is dose dependent, but with the 1764/27- virus, the peak of ICP22/47 expression is at an MOI of 0.5. No expression of ICP22
25 was detected at any MOI from the 1764/27-/4- virus. This phenomenon was investigated further and found to also apply to exogenous genes, for example CMV driven GFP, inserted into the 1764/27- virus but not the less disabled 17+/27- virus or the more disabled 1764/27-/4- virus. Virus strains 1764/27-/4-pR19GFP and 1764/27-/4-pR20.5 demonstrate a dose dependent pattern of GFP expression.
30 However, with virus strain 1764/27-/pR20.5 the pattern of GFP expression follows that of ICP22/47, with expression levels peaking at an MOI of 0.5. The reason for this expression pattern is unclear, but a similar dose effect has been reported

previously when ICP0 and ICP4 were used in combination to activate IE gene target promoters in transient transfection assays (Gelman and Silverstein 1987). Whatever the mechanism, a linearly dose dependent pattern of transgene depression such as is directed by the most disabled virus is more desirable in gene delivery applications
5 than the pattern shown by the less disabled virus.

Example 7: Virus strain 1764/27-/4- directs high efficiency gene delivery to cultured neurons.

In order to test the ability of 1764 27- 4- viruses to deliver exogenous genes
10 to cultured neurons, primary cultures of DRG neurons were infected at an MOI of 10 with 1764/27-/4-/pR20.5. Mock infected neurons or neurons infected with virus strain 17+/27-/pR19GFP were used as controls. One week post infection, neurons were photographed under phase contrast and fluorescence microscopy. Both viruses were able to efficiently deliver exogenous genes to neurons in culture. However, by
15 7 days post infection, the 17+/27- virus caused considerable toxic effects, with loss of supporting cells, clumping of neuronal cell bodies and retraction of neuronal processes. In contrast, cells infected the 1764/27-/4- virus are indistinguishable in morphology from mock infected cells under phase contrast microscopy. Neurons infected with the 1764/27-/4- virus express abundant levels of GFP, in both neuronal
20 cell bodies and processes, demonstrating efficient gene delivery.

Example 8: Virus strain 1764/27-/4- directs the expression of delivered genes for extended periods in neuronal and non-neuronal cells.

In order to test whether the 1764/27-/4- virus could establish a persistent
25 infection, Vero cells at 50% confluence were infected at an MOI of 10 with virus strain 1764/27-/4-/pR20.5. Following infection, cells were maintained in 2% serum at 34°C, and reporter gene expression was monitored at 3, 7 and 23 days post infection. At the last time point (23 days), a duplicate well was superinfected at an MOI of 5 with virus strain 17+/27- (expressing no reporter gene). GFP expression
30 was evident at all time points during the experiment, and that unlike in the previously published work (Samaniego *et al.* 1998), superinfection is not necessary for transgene expression to occur. GFP expression is present in reducing numbers of

cells over time, as in previous work, which is probably due to cell division in the culture and the finite life span of both transduced and untransduced cells.

Superinfection of the cultures at 23 days does increase GFP expression levels slightly but the effect is not marked. This demonstrates that the genome of virus strain

- 5 1764/27-/4-/pR20.5 is capable of persisting in the nucleus of infected cells, indicating the low toxicity of the vector. At all times up to the end of the experiment, the cells infected with virus strain 1764/27-/4-/pR20.5 were indistinguishable from mock infected cells.

- In order to extend these findings, the expression characteristics of virus strain
10 1764/27-/4-/pR20.5 in rat organotypic hippocampal slice cultures was also investigated. Organotypic hippocampal slice cultures were prepared and infected 7 days later with 1×10^6 pfu of 1764/27-/4-/pR20.5. Infected slices were stained for lacZ together with expression of GFP at 2 days, 7 days and 21 days after infection.

- A high level of transgene expression was observed at 2 days post infection.
15 was maintained at 1 week and was still clearly detectable in the cultures 3 weeks after infection, albeit in a reduced number of cells. The processes of transduced neurons, even at the 3 week time point, were not swollen or beaded suggesting that degeneration of transduced neurons had not occurred. Infected cultures maintained normal morphology and did not differ from mock-infected control cultures at any
20 time. These results again suggest that the toxic effects of 1764 27- 4-pR20.5 on transduced cells are minimal, at least by morphological criteria, even over extended periods in culture.

References

- Fink *et al.* 1996 Ann Rev Neurosci 19; 265-287
- Goldstein and Weller 1988 J Virol 62; 196-205
- 5 Ho and Mokarski 1988 Virology 167; 279-283
- Lokensgard *et al.* 1994 J. Virol 68; 7148-7158.
- Dobson *et al.* 1990 J. Virol. 69; 2264-2270
- Goins *et al.* 1994 J. Virol 68; 2239-2252.
- Lachmann and Efsthathiou 1997 J. Virol 71; 3197-3207
- 10 Samaniego *et al.* 1998 J. Virol 72; 3307-3320
- Krisky *et al.* 1998 Gene Therapy 5; 1593-1603
- Thomas *et al.* 1999 J. Virol 73; 7399-7409
- Howard *et al.* 1998 Gene Therapy 5; 1137-1147
- Wagstaff *et al.* 1998 Gene Therapy 5; 1566-1570
- 15 Samaniego LA *et al.* J.*Virol. (1995); 69: 5705-5715
- Coffin RS and Latchman DS (1996). In: Genetic Manipulation of the Nervous System (DS Latchman Ed.) pp 99-114: Academic Press, London.
- Coffin RS, *et al.* (1996) Gene Therapy, 3, 886-891.
- Ace CI, *et al.* (1989) J. Virol., 63, 2260-2269.
- 20 Smith, IL *et al.*, 1992, Virology 186: 74-86.
- Rice, SA and Knipe DM., 1990, J. Virol 64: 1704-1715.
- DeLuca NA *et al.* J. Virol. 1985; 56: 558-570.
- MacFarlane M, *et al.* (1992) J. Gen. Virol., 73, 285-292.
- Lokensgard JR, *et al.* (1994) J. Virol., 68, 7148-7158.
- 25 Gossen M and Bujard H, 1992, PNAS 89: 5547-5551.
- Gossen M *et al.*, 1995, Science 268: 1766-1769.
- Smiley, J. R., and J. Duncan. 1997. J. Virol. 71: 6191-6193.
- Johnson *et al.* 1994. J. Virol 68; 6347-6362.
- Johnson *et al.* 1992. J. Virol; 2952-2965
- 30 Wu *et al.* 1996. J. Virol 70; 6358-6369
- Sacks and Shaffer 1987 J Virol; 829-839
- Stow and Stow 1986 J Gen Vir 67; 2571-2585

- Post and Roizman 1981 Cell 25; 227-232
- Sears *et al.* 1985 J. Virol 55; 338-346
- Deluca and Schaffer 1987 NAR 15; 4491-4511
- Samaniego *et al.* 1995 J. Virol 69; 5705-5715
- 5 Samaniego *et al.* 1997 J. Virol 71; 4614-4625
- Batterson and Roizman 1983 J Virol 46; 371-377
- Pellet *et al.* 1985 PNAS 82; 5870-5874
- Bloom *et al.* 1995 Mol Br Res 3 1; 48-60
- Carpenter and Stevens 1996 Hum Gen Ther 7; 1447-1454
- 10 Lokensgard *et al.* 1997 J. Virol 71; 6714-6719
- Preston and Nicholl 1997 J Viol 71; 7807-7813
- Gelmen and Silverstein 1987 J Virol 61; 2286-2296 and 3167-3172
- Sze and Hermen 1992 Virus Res. 26; 141-152
- Thompson *et al.* 1998 Virus Genes 1(3); 275-286
- 15 Meignier *et al.* 1988 J. Infect. Dis. 159; 602-614

CLAIMS

1. Use of a replication incompetent herpes virus comprising:
 - 5 (a) a mutation which prevents or reduces the expression of at least two immediate early genes; and
 - (b) a heterologous gene operably linked to a promoter active during herpes virus latencyin the manufacture of a medicament for use in treating or preventing a central
10 nervous system disorder.
2. A use according to claim 1 wherein said virus is a herpes simplex virus 1.
- 15 3. A use according to claim 1 wherein said virus is a herpes simplex virus 2.
4. A use according to any one of the preceding claims, wherein said virus lacks a functional VMW65 gene due to a mutation in said gene which abolishes
20 its transcriptional-activation activity.
5. A use according to any one of the preceding claims, wherein said immediate early genes are selected from genes encoding ICP0, ICP4, ICP22 and ICP27.
25
6. A use according to any one of the preceding claims, wherein said virus lacks a functional gene encoding ICP4 and a functional gene encoding ICP27 and which has an inactivating mutation in the gene encoding vmw65 abolishing its transcriptional-activation activity.
30
7. A use according to any one of the preceding claims, wherein said virus lacks functional genes encoding ICP27, ICP4, ICP0 and ICP22.

8. A use according to any one of the preceding claims, wherein said promoter is chimeric promoter containing elements of a latency associated transcript (LAT) region of a herpes virus and a non-LAT promoter.

5

9. A use according to any preceding claim wherein said promoter is or contains elements from the LAP2 HSV promoter.

10. A use according to anyone of the preceding claims wherein said promoter contains non-HSV elements such as an internal ribosome entry site (IRES) sequence or other sequence capable of performing a similar function.

11. A use according to any one of the preceding claims wherein said heterologous gene encodes a polypeptide of therapeutic use.

15

12. A use according to claim 11 wherein said polypeptide is capable of preventing or slowing neuronal degeneration, reversing neuronal degeneration or stimulating cells remaining in the central nervous system of a subject suffering from a neurodegenerative disorder to perform a function of cells absent in said subject.

20

13. A use according to any one of the preceding claims wherein said medicament is for delivery of a gene to two or more connected sites in the central nervous system by administration to a single site in the nervous system.

14. A use according to any one of the preceding claims wherein said medicament is for injection into the striatum or the ventricles within the brain.

15. A method of determining whether a gene has an effect on a phenotype associated with a central nervous system disorder or on a cell of the central nervous system which is relevant to a central nervous system disorder, which method comprises:

30

(i) inoculating into a cell of the central nervous system with a replication incompetent herpes virus comprising:

(a) a mutation which prevents or reduces the expression of at least two immediate early genes; and

5 (b) a heterologous gene operably linked to a promoter active during herpes virus latency; and

(ii) monitoring a phenotype of said disorder or an effect of expression of said gene on said cell to determine thereby whether said gene has an effect on said cell or said phenotype.

10

16. A method according to claim 15 wherein said cell is inoculated *in vivo*.

17. A method according to claim 15 or 16 wherein said virus is as defined
15 in any one of claims 2 to 10.

18. A method according to any one of claims 15 to 17, wherein said gene is selected from a gene encoding a neurotropic factor, an enzyme, a protein such as Sonic Hedgehog or Nurr-1 and a gene deficient in a metabolic disease.

20

19. A method according to any one of claims 15 to 18, wherein said gene is implicated in a central nervous system disorder.

20. A method according to any one of claims 15 to 19, wherein said
25 disorder is a neurodegenerative disorder.

21. A method according to any one of claims 15 to 19, wherein said disorder is selected from stroke, Parkinson's disease, Alzheimer's disease, Tay Sachs disease and a mucopolysaccharide disease.

30

22. A method according to any one of claims 15 to 21, wherein said cell is a mammalian cell.

23. A method according to any one of claims 15 to 22, wherein said cell is a neuron.

5 24. A method according to any one of claims 15 to 23, wherein said cell is in a non-human animal which models said disorder.

25. A method according to claim 24 wherein said animal is a transgenic animal.
10

26. A method according to any preceding claim wherein said phenotype is selected from a motor control disorder and a cognitive deficit.

27. A method according to any one of claims 15 to 26 wherein said
15 heterologous gene is expressed in said cell following retrograde transport along axonal processes from the site of contact between said cell and said virus.

28. A method according to any one of claims 15 to 27 wherein said heterologous gene is expressed in at the site of contact between said cell and said
20 virus and at least one other site in the central nervous system which site is distinct from the site of contact.

29. A method of screening genes implicated in a central nervous system disorder to identify a target for gene therapy or for small molecule
25 modulators, which method comprises a method according to any one of the preceding claims.

30. A method of treating a subject suffering from a disorder of the central nervous system, or of preventing a disorder of the central nervous system in a subject
30 at risk thereof, which method comprises administering a therapeutically effective amount of a replication incompetent herpes virus comprising:

(a) a mutation which prevents or reduces the expression of at least two immediate early genes; and

(b) a heterologous gene operably linked to a promoter active during herpes virus latency

5 to a subject suffering from a disorder of central nervous system.

31. A method according to claim 30 wherein said virus is administered to at least one site in the central nervous system and the gene is delivered to at least one connected site within the central nervous system.

10

32. A method according to any one of claims 30 to 31 wherein said virus is as defined in any one of claims 2 to 10.

33. A method according to any one of claims 30 to 32 wherein said
15 heterologous gene encodes a polypeptide or antisense RNA of therapeutic use.

34. A method according to claim 33 wherein said polypeptide or RNA is capable of preventing or slowing neuronal degeneration, reversing neuronal degeneration or stimulating cells remaining in the central nervous system of a subject
20 suffering from a neurodegenerative disorder to perform a function of cells absent in said subject.

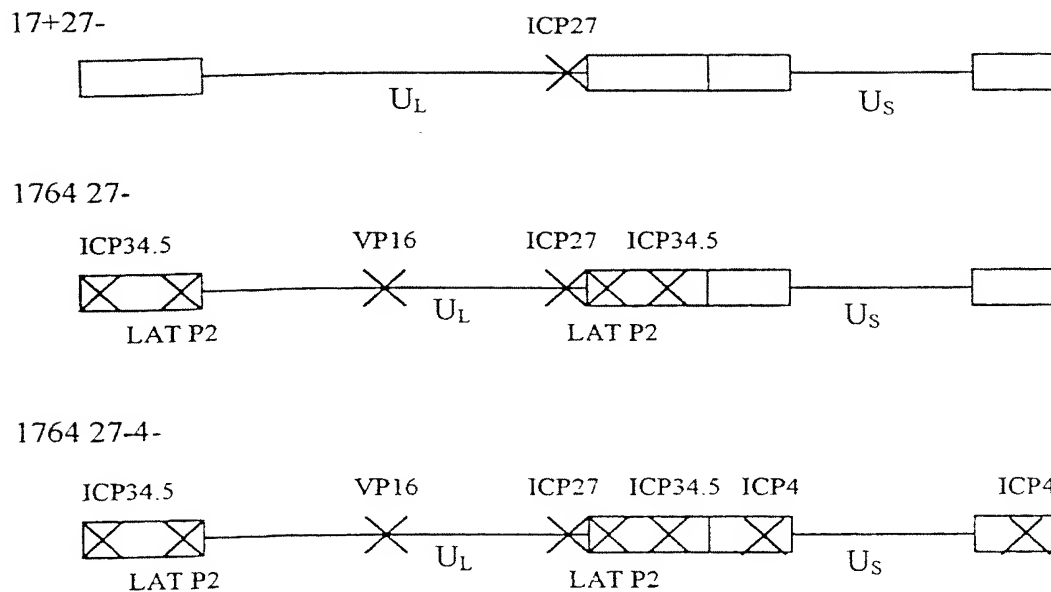
35. A method according to any one of claims 30 to 34 wherein said virus is administered to said subject by injection into the brain.

25

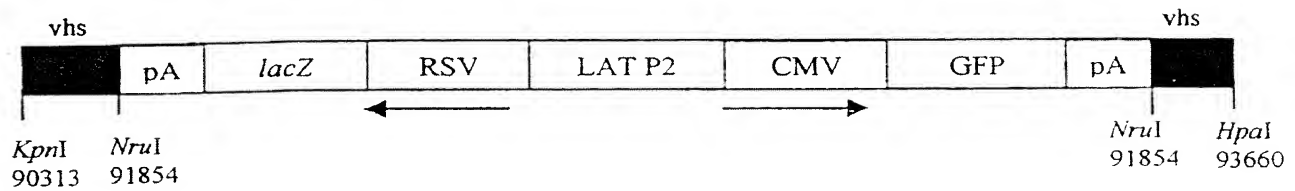
36. An agent for treating or preventing a disorder of the central nervous system comprising a replication incompetent herpes virus comprising:

(a) a mutation which prevents or reduces the expression of at least two immediate early genes; and

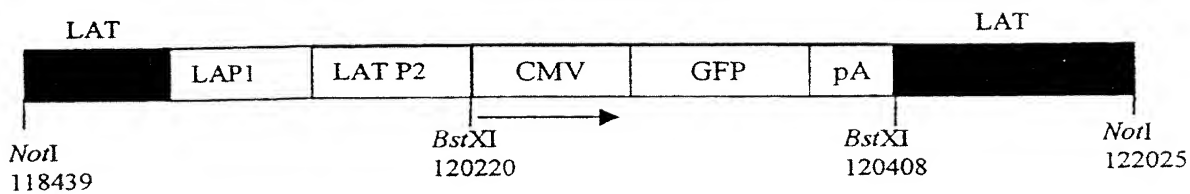
30 (b) a heterologous gene operably linked to a promoter active during herpes virus latency.

Figure 1**a) Vector backbones****b) Promoter cassettes**

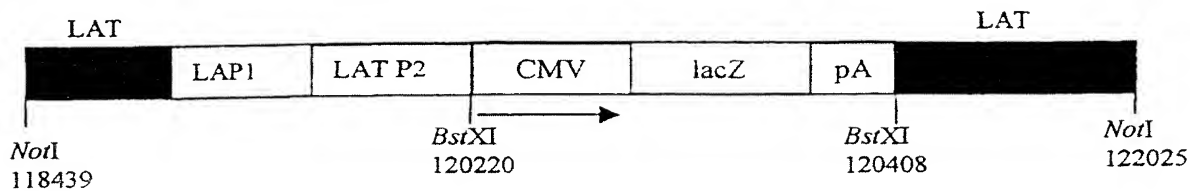
pR20.5 (insertion site = vhs)



pR19GFP (insertion site = LAT region)



pR19lacZ



INTERNATIONAL SEARCH REPORT

International Application No

PC1/GB 00/04981

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/86 A61K48/00 G01N33/50 A61P25/00 A61P25/16
A61P25/28

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

WPI Data, PAJ, EPO-Internal, BIOSIS, MEDLINE, CHEM ABS Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 98 30707 A (UNIV LONDON ;COFFIN ROBERT STUART (GB); LATCHMAN DAVID SEYMOUR (GB) 16 July 1998 (1998-07-16) cited in the application page 1, line 4 - line 8 page 1, line 11 -page 2, line 22 page 2, line 25 -page 3, line 3 page 3, line 4 - line 14 page 13, line 6 -page 14, line 3 page 9, line 11 - line 30 page 14 -page 18; examples 1-5,8 --- -/--	1-5,8,9, 11-36
Y		6,7

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
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Date of the actual completion of the international search

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INTERNATIONAL SEARCH REPORT

International Application No

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>WO 97 20935 A (EFSTATHIOU STACEY ;UNIV CAMBRIDGE TECH (GB); LACHMANN ROBIN HENRY) 12 June 1997 (1997-06-12) page 1, line 2 - line 12 page 6, line 24 -page 8, line 18 page 11, line 13 -page 13, line 5 page 16, line 20 -page 17, line 23 page 24, line 11 -page 25, line 24</p> <p>----</p>	1-3,5, 10-13, 30-36
X	<p>WO 99 60145 A (NEURO VEX LIMITED ;COFFIN ROBERT STUART (GB); LATCHMAN DAVID SEYMO) 25 November 1999 (1999-11-25)</p> <p>page 1, line 5 - line 9 page 2, line 17 -page 3, line 14 page 3, line 19 -page 4, line 27 page 5, line 10 - line 22 page 7, line 3 -page 9, line 4</p> <p>----</p>	1-3,5,8, 9,11-13, 15-23, 26-36
Y	<p>THOMAS S K ET AL: "Equine herpesvirus 1 gene 12 can substitute for vmw65 in the growth of herpes simplex virus (HSV) type 1, allowing the generation of optimized cell lines for the propagation of HSV vectors with multiple immediate-early gene defects." JOURNAL OF VIROLOGY, vol. 73, no. 9, 1999, pages 7399-7409, XP002167072 ISSN: 0022-538X cited in the application page 7399, paragraph 1 -page 7400, paragraph 3</p> <p>----</p>	6,7
A	<p>LACHMANN R H ET AL: "THE USE OF HERPES SIMPLEX VIRUS-BASED VECTORS FOR GENE DELIVERY TO THE NERVOUS SYSTEM" MOLECULAR MEDICINE TODAY,GB,ELSEVIER, CAMBRIDGE, September 1997 (1997-09), pages 404-411, XP000783616 ISSN: 1357-4310 the whole document</p> <p>-----</p>	

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/GB 00/04981

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9830707 A	16-07-1998	AU 5566998 A BR 9806866 A CN 1250480 T EP 1021553 A	03-08-1998 18-04-2000 12-04-2000 26-07-2000
WO 9720935 A	12-06-1997	AU 712793 B AU 1102997 A CA 2238981 A EP 0865493 A JP 2000501933 T US 6193980 B	18-11-1999 27-06-1997 12-06-1997 23-09-1998 22-02-2000 27-02-2001
WO 9960145 A	25-11-1999	AU 3946699 A EP 1080215 A	06-12-1999 07-03-2001